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Synthesis of calix[4]arene podands bearing two and four histidine or glycine groups at the lower rim. Complexation properties towards cobalt(II) chloride

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Abstract

Four calix[4]arene podands displaying two or four glycine or histidine arms at the lower rim have been synthesised and fully characterised. Their coordinating behaviour towards cobalt(II) chloride has been investigated. © 1999 Elsevier Science Ltd. All rights reserved.

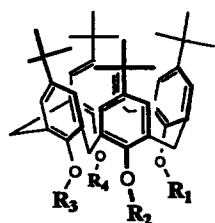
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In the field of supramolecular chemistry, the calix[4]arene platform **1** displays interesting organising properties, notably, as recently reviewed by Matt and co-workers¹ for the building of ester- or amide-containing ligands for alkaline and alkaline-earth cations and heterocycle-containing ones for transition metal species. Surprisingly, the chelating behaviour of amino acid pendant arms grafted on such macrocycles has not been investigated until now. Nevertheless, the introduction of amino acids at the upper and lower rims of the calix[4]arene platform has been described, leading to water-soluble chiral entities. At the upper rim, Shinkai and co-workers² introduced four L-cysteine subunits, while more recently, Ungaro and co-workers³ described the selective introduction of two and four alanine groups. At the lower rim, Warner and co-workers⁴ grafted, after isolation of the corresponding succinimidoyl activated ester, four alanine units separated from the calixarene platform by an *O*-acetyl link. The resulting chiral podand was used in this case as mobile phase additive in capillary electrophoresis.

As part of our general project dedicated to the building of lipophilic ligands including heterocyclic chelating arms preorganised on a calixarene platform,⁵ we attempted to prepare transition metal complexing agents bearing active amino acid subunits which could mimic the activity of some metallo-enzymes. Thus, in parallel to investigations related to the synthesis of a carbonic anhydrase biomimetic, we attempted to prepare a tetra-histidyl calix[4]arene podand in the cone conformation. Our first experiments

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involved coupling reaction between the tetra-acid chloride **4**,⁶ obtained by direct chlorination of the tetra-acid **3**⁶ with SOCl_2 , and histidine methyl ester or, in the presence of DCC and HOBT, between **3** and *N*-benzyl histidine methyl ester. In both cases, the resulting products were in fact mixtures of intermediates we were unable to separate correctly.

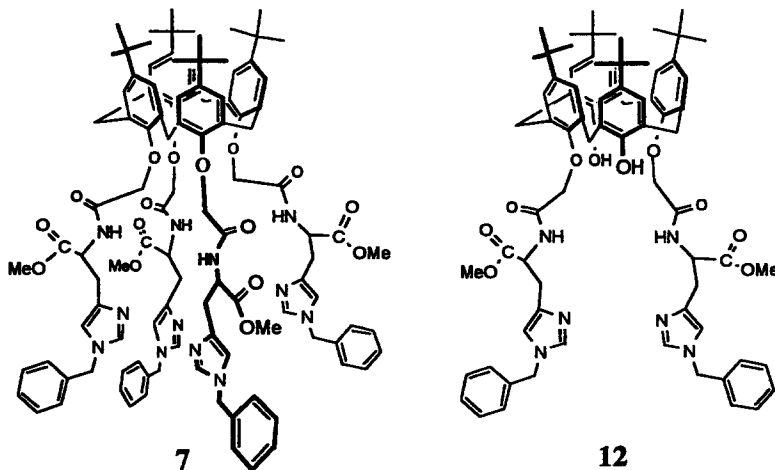


1:	$\text{R}_1 = \text{R}_2 = \text{R}_3 = \text{R}_4 = \text{H}$	8:	$\text{R}_1 = \text{R}_3 = \text{H}; \text{R}_2 = \text{R}_4 = \text{CH}_2\text{COOEt}$
2:	CH_2COOEt	9:	$\text{H}; \text{CH}_2\text{COOH}$
3:	CH_2COOH	10:	$\text{H}; \text{CH}_2\text{COO-Nsuccinimide}$
4:	CH_2COCl	11:	$\text{H}; \text{CH}_2\text{CONHCH}_2\text{COOEt}$
5:	$\text{CH}_2\text{COO-Nsuccinimide}$		
6:	$\text{CH}_2\text{CONHCH}_2\text{COOEt}$		

Adapting the DCC/NHS procedure of Warner,⁴ which involves the isolation of the tetra-succinimidoyl activated ester **5**, to the synthesis of the model compound tetra-glycyl **6**, did not give reproducible results. The yield of 70% given for **5** was obtained with a longer reaction time (48 h instead of 12 h at 25°C) and after a careful washing of the DCU precipitate. The quadruple peptide-bound formation gave reproducible results but with a longer reaction time (72–120 h instead of 24 h); **6** was thus obtained pure in 60% yield after chromatography.⁷

In parallel, the tetra-*N*-benzyl histidine methyl ester derivative **7** was obtained pure with a yield of 40%; nevertheless, the washing step resulted in the formation of a milky emulsion between CH_2Cl_2 and H_2O , which necessitated decantation times of ca. 72 h. The presence of polluting residual DCU also necessitated a purification by chromatography (Al_2O_3 , CH_2Cl_2 :MeOH 98:2).⁷

The versatile organising properties of the calix[4]arene platform were also tested with the introduction of two amino acid residues. That was done by using the same synthetic procedure, but without protecting the two residual OH groups located at the 26th and the 28th positions. The bisactivated ester **10** was thus obtained with a yield of 75% as a white solid. Reaction of **10** with GlyOEt or Hist(Bzl)OMe at 25°C during 5 days under anhydrous conditions afforded the bisglycyl and bishistidyl podands **11** and **12**, with respective yields of 65 and 52%, after chromatography.⁷



¹H NMR analysis of **12** showed that the chirality of the histidyl arms induces a discrimination of the OCH_2 protons which appear as an AB system at 4.15–5.00 ppm ($J_{AB}=14.7$ Hz); this phenomenon is also observed, but at a lesser degree, with **7**, in which the eight OCH_2 protons appear as a strong AB system at

4.57–4.65 ppm ($J_{AB}=14.7$ Hz), overloaded by the low field component of the Ar-CH₂-AR AB resonance signal. As expected, no differentiation was observed with the glycylic analogues **6** and **11**.

Thus, the presence of histidyl groups at the lower rim of the calixarene platform generates a prohelical structure which could be strengthened in **12** by hydrogen bonding between the free OH groups and the amide carbonyl subunits; nevertheless, the expected shift of ca. 50 cm⁻¹ for the corresponding IR band is not observed. However, the higher space available for the two histidyl groups in **12**, with regard to **7**, explains the expanded AB system observed for the OCH₂ protons.

Attempts to prepare complexes of Co(II) with the glycylic podands **6** and **11** were unsuccessful, but both histidinyl analogues **7** and **12** formed blue-coloured species **13** and **14**, with a tendency to give crystals unfortunately unsuitable for X-ray analysis. UV spectroscopy notably showed the presence of a metal-centred absorption band at 615 nm which was used to determine the corresponding stoichiometries via Job plot experiments. These were carried out in CH₂Cl₂/EtOH and showed that **13** displays an exact M₂L stoichiometry, while **12** gave relatively ambiguous results leading to an M_{1.5}L (M₃L₂). Positive mode ES-MS analyses confirmed the M₂L nature of **13** with, notably, the molecular peak at 2069.7 amu ([7+2 CoCl₂-Cl]⁺). The M₃L₂ stoichiometry of the isolated complex **14** was not evidenced by this technique, which provides two major peaks at 1340.4 amu ([12+CoCl₂-Cl]⁺), and 1247.6 amu ([12+H]⁺). Elemental analysis of **14** was consistent with the presence of one cobalt cation, and UV-visible spectroscopy of the isolated complex showed that at 615 nm, with regards to **13**, its stoichiometry is of the ML type. IR analysis (KBr pellets) showed that the complexation of Co(II) should not involve the carbonyl groups, no specific shift of the corresponding elongation bands being observed between the free ligands **7** and **12** and their complexes. A participation of the amide N-H subunits to the complexation process could be related, although ambiguously, to the apparition of a relatively intense band at 1519.0 cm⁻¹ (N-H bending).

The grafting of amino acid residues on a calix[4]arene platform lead to potent transition metal complexing agents which could display metallo-enzyme biomimetic activities. The preparation of a tris-histidinyl podand designed to mimic the carbonic anhydrase active centre is now in progress.

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References

1. Wieser, C.; Dieleman, D. B.; Matt, D. *Coord. Chem. Rev.* **1997**, *165*, 93–161.
2. Nagasaki, T.; Tajiri, U.; Shinkai, S. *Rec. Trav. Chim. Pays Bas* **1993**, *112*, 407–411.
3. Sansone, F.; Barbosa, S.; Casnati, A.; Pochini, A.; Ugozzoli, F.; Ungaro, R. *Eur. J. Org. Chem.* **1998**, 897–905.
4. Sanchez Peña, M.; Zhang, Y.; Thibodeaux, S.; McLaughlin, M. L.; Munoz de la Peña, A.; Warner, I. M. *Tetrahedron Lett.* **1996**, *37*, 5841–5844.
5. (a) Regnouf-de-Vains, J.-B.; Lamartine, R. *Helv. Chim. Acta* **1994**, *77*, 1817–1825. (b) Regnouf-de-Vains, J.-B.; Pellet-Rostaing, S.; Lamartine, R. *Tetrahedron Lett.* **1995**, *36*, 5745–5748. (c) Regnouf-de-Vains, J.-B.; Lamartine, R.; Fenet, B.; Bavoux, C.; Thozet, A.; Perrin, M. *Helv. Chim. Acta* **1995**, *78*, 1607–1619. (d) Pellet-Rostaing, S.; Regnouf-de-Vains, J.-B.; Lamartine, R. *Tetrahedron Lett.* **1996**, *37*, 5889–5892. (e) Pellet-Rostaing, S.; Regnouf-de-Vains, J.-B.; Lamartine, R.; Meallier, P.; Guittonneau, S.; Fenet, B. *Helv. Chim. Acta* **1997**, *80*, 1229–1243. (f) Regnouf-de-Vains, J.-B.; Lamartine, R.; Fenet, B. *Helv. Chim. Acta* **1998**, *81*, 661–669. (g) Regnouf-de-Vains, J.-B.; Lamartine, R. *Tetrahedron Lett.* **1996**, *37*, 6311–6314.

6. The tetra-ester **2**, tetra-acid **3**, diester **8**, and diacid **9** were synthesised according to the literature: (a) Iwamoto, K.; Shinkai, S. *J. Org. Chem.* **1992**, *57*, 7066–7073. (b) Cang, S. K.; Cho, I. *J. Chem. Soc., Perkin Trans. 1* **1986**, 211–214. (c) Arnaud-Neu, F.; Collins, E. M.; Deasy, M.; Ferguson, G.; Harris, S. J.; Kaiyner, B.; Lough, A. J.; Mc Kervey, M. A.; Marques, E.; Ruhl, B. L.; Schwing-Weill, M.-J.; Seward, E. M. *J. Am. Chem. Soc.* **1989**, *111*, 8681–8691. The acid chloride **4**: Arnaud-Neu, F.; Barret, G.; Cremin, S.; Deasy, M.; Ferguson, G.; Harris, S. J.; Lough, A. J.; Guerra, L.; Mc Kervey, M. A.; Schwing-Weill, M.-J.; Schwinte, P. *J. Chem. Soc., Perkin Trans. 2* **1992**, 1119–1124.
7. General: Melting points (°C, uncorrected) were determined on an electrothermal 9100 capillary apparatus. ¹H and ¹³C NMR spectra were recorded on a Bruker AM 300 (300.13 and 75 MHz) or DRX 500 (500.13 and 125.75 MHz), (CDCl₃, TMS as internal standard, chemical shifts in ppm, J in hertz). Mass spectra (electrospray, ES) were recorded on a Platform Micromass apparatus at the Service Central d'Analyse du CNRS, Solaize. Infra-red was performed on a Mattson 5000 FT apparatus (ν in cm⁻¹) and UV spectra were recorded on a Shimadzu UV 2401 PC apparatus, λ_{\max} in nm, ϵ in dm³ mol⁻¹ cm⁻¹. Elemental analyses were performed at the Service Central de Microanalyse du CNRS, Solaize. Macherey-Nagel TLC plates were used for chromatography analysis (SiO₂, Polygram SIL G/UV254, Ref. 805021). All commercially available products were used without further purification unless otherwise specified. Compound **6**: The tetra-acid **3** (0.88 g, 1 mmol) was added to a solution of *N*-hydroxysuccinimide (0.48 g, 4.2 mmol) in dry ethyl acetate. A solution of DCC (0.87 g, 4.2 mmol) in dry ethyl acetate was added and stirred 48 h at 25°C under anhydrous conditions. The resulting precipitate was filtered and washed three times with ethyl acetate and the filtrate was concentrated in vacuo to give **5** in 70% yield as white product. The calixarene **5** (0.55 g, 0.39 mmol) was suspended in dry CH₂Cl₂. A solution of GlyOEt, HCl (0.24 g, 1.73 mmol) and diisopropylethylamine (0.79 g, 6.1 mmol) in dry CH₂Cl₂ was added and the mixture was stirred at 25°C for 5 days. After addition of H₂O, the product was extracted with CH₂Cl₂. The combined organic phases were washed with water and evaporated in vacuo. The mixture was chromatographed (Al₂O₃, CH₂Cl₂:MeOH 98:2) to give **6** (0.28 g, 60%) as white product. M.p.: 193.2°C. IR (KBr): 1663.8 (CO(NH)); 1751.1 (CO(O)); 3329.3 (NH). UV (CH₂Cl₂): 274.5 (3300). ¹H NMR (CDCl₃): 1.09 (s, 36H, Me₃C); 1.28 (t, J=7.35, 12H, CH₃CH₂); 3.31, 4.56 (AB, J_{AB}=13.3, 8H, Ar-CH₂Ar); 4.12 (d, J=5.89, 8H, N-CH₂); 4.16 (q, J=7.36, 8H, CH₃CH₂); 4.58 (s, 8H, OCH₂); 6.82 (s, 8H, H-Ar); 7.92 (t, 4H, NH). ¹³C NMR: 14.12 (CH₃CH₂); 31.12 (Ar-CH₂-Ar); 31.31 (Me₃C); 33.92 (Me₃C); 40.99 (CH₂-NH); 61.31 (CH₃CH₂); 74.18 (OCH₂); 125.96 (3,5-Ar); 132.66, 146.14, 152.49 (1,2,4,6-Ar); 170.11 (CONH); 170.37 (COOEt). ES-MS (pos. mode): 1243.9 [6+Na]⁺. Anal. calcd for C₆₈H₉₂O₁₆N₄ (1220): C 66.89; H 7.59; N 4.59; O 20.96; found: C 66.54; H 7.75; N 4.14; O 20.59. Compound **7**: Same procedure, from Hist(Bzl)OMe, 2HCl (0.63g, 1.73 mmol); (**7**: 0.29 g, 40%, white powder). M.p. 189.3°C. IR: 1671.2 (CO(NH)); 1743.5 (CO(O)); 3316 (NH). UV (CH₂Cl₂): 282.0 (3380). ¹H NMR: 1.05 (s, 36H, Me₃C); 3.03 (br m, 8H of CH₂-His and 4H of ArCH₂Ar); 3.54 (s, 12H, OCH₃); 4.57, 4.65 (AB, J_{AB}=14.7, 8H, OCH₂); 4.61 (d, J_{AB}=14.7, 4H, ArCH₂Ar); 4.88 (dt, J₁=7.35, J₂=6.61, 4H-C_αHis); 4.98 (s, 8H, CH₂-Ph); 6.69 (s, 4H-Ar); 6.71 (s, 4H, H₍₂₎Im); 7.10 (d, J=5.15, 8H_(o)-Ph); 7.28 (m, 12H_(m,p)-Ph); 7.39 (s, 4H₍₄₎Im); 8.28 (d, J=8.1, 4 NH). ¹³C NMR: 30.62 (CH₂-Im); 31.79 (Me₃C); 32.32 (Ar-CH₂-Ar); 34.22 (Me₃C); 51.09 (CH₂-Ph); 52.57 (OMe); 52.83 (C_αHis); 74.19 (OCH₂); 117.53 (4-Im); 125.68, 126.02 (2-Im); 127.65, 128.54, 129.32 (2,3,4,5,6-Ph); 136.80 (2,6-Ar); 137.31 (3,5-Ar); 138.6 (4-Ar, 1-Ph); 145.37 (1-Ar); 153.73 (5-Im); 170.49 (CONH); 172.71 (COOMe). ES-MS (pos. mode): 1869.1 [7+Na]⁺, 934.8 [7+H+Na]²⁺/2, 923.7 [7+2 H]²⁺/2, 1847.1 [7+H]⁺. Anal. calcd for C₁₀₈H₁₂₄O₁₆N₁₂ (1846.26): C 70.26, H 6.77, N 9.10, O 13.87; found: C 69.96, H 6.96, N 8.99, O 14.33. Compound **10**: Prepared according to **5** (see above). M.p.: 189°C. ¹H NMR (CDCl₃): 0.91 (s, 18H, Me₃C); 1.29 (s, 18H, Me₃C); 2.81 (s, 8H, CH₂CH₂); 3.37, 4.36 (AB, J_{AB}=13.6, 8H, Ar-CH₂Ar); 5.20 (s, 4H, OCH₂); 6.58 (s, 2H, OH); 6.72 (s, 4H, H-Ar); 7.06 (s, 4H, H-Ar). Compound **11**: The diacid **9** (0.77 g, 1 mmol) was added to a solution of *N*-hydroxysuccinimide (0.25 g, 2.2 mmol) in dry ethyl acetate. A solution of DCC (0.47 g, 2.2 mmol) in dry ethyl acetate was added and stirred 48 h at 25°C under anhydrous conditions. The resulting precipitate was filtered and washed three times with ethyl acetate and the filtrate was concentrated in vacuo to give **10** in 75% yield as a white product. The calixarene **10** (0.60 g, 0.63 mmol) was suspended in dry CH₂Cl₂. A solution of GlyOEt, HCl (0.19 g, 1.38 mmol) and diisopropylethylamine (0.49 g, 3.8 mmol) in dry CH₂Cl₂ was added and the mixture was stirred at 25°C for 5 days. After addition of H₂O, the product was extracted with CH₂Cl₂. The combined organic phases were washed with water and evaporated in vacuo. The mixture was chromatographed (Al₂O₃, CH₂Cl₂:MeOH 98.5:1.5) to give **11** (0.39 g, 65% yield) as a white product. M.p.: 150°C. IR (KBr): 1691.3 (CO(NH)); 1752.2 (CO(O)); 3324.7 (NH). UV (CH₂Cl₂): 289.0 (6700). ¹H NMR (CDCl₃): 1.06 (s, 18H, Me₃C); 1.22–1.27 (m, 24H, Me₃C+CH₃CH₂); 3.43, 4.23 (AB, J_{AB}=13.2, 8H, Ar-CH₂Ar); 4.11–4.25 (m, 10H, CH₃CH₂+NCH₂); 4.63 (s, 4H, OCH₂); 6.94 (s, 4H, H-Ar); 7.09 (s, 4H, H-Ar); 7.92 (s, 2H, OH); 9.30 (t, J=5.15, 2H, NH). ¹³C NMR: 14.06 (CH₃CH₂); 30.99, 31.61 (Me₃C); 32.18 (Ar-CH₂-Ar); 33.90, 34.13 (Me₃C); 41.31 (CH₂NH); 61.39 (CH₃CH₂); 74.83 (OCH₂); 125.51, 126.20 (3,5-Ar); 127.08, 132.44, 142.83, 148.21, 149.37, 149.75 (1,2,4,6-Ar); 169.00 (CONH); 169.35 (COOEt). ES-MS (pos. mode): 957.6 [11+Na]⁺. Anal. calcd for C₅₆H₇₄O₁₀N₂+H₂O (952): C 70.11; H 8.22; N 3.29; O 18.38; found: C 70.56; H 8.03; N 2.95; O 18.46. Compound **12**: Same procedure, from Hist(Bzl)OMe, 2HCl (0.50 g, 1.38 mmol); (**12**:

0.41 g, 52%, white powder). M.p. 199.5°C. IR (KBr): 1673.6 (CO(NH)); 1749.6 (CO(O)); 3322.6 (NH). UV (CH₂Cl₂): 290.0 (7400). ¹H NMR: 1.02 (s, 18H, Me₃C); 1.25 (s, 18H, Me₃C); 3.10, 3.18 (ABd, J_{AB}=14.0, J=5, 4H, CH₂-His); 3.11, 4.19 (AB, J_{AB}=13, 4H, Ar-CH₂-Ar); 3.35, 4.03 (AB, J_{AB}=14, 4H, Ar-CH₂-Ar); 3.65 (s, 6H, OCH₃); 4.15, 5.00 (AB, J_{AB}=14.7, 4H, OCH₂); 4.25, 4.53 (AB, J_{AB}=15.4, 4H, CH₂Ph); 5.00 (sext, J₁=7.4, J₂=5, 2H-C_αHis); 6.54 (d, J=0.5, 2H₍₄₎Im); 6.86 (AB, J_{AB}=2, 4H-Ar); 6.92 (dd, J₁=7.7, J₂=2, 4H_(o)-Ph); 7.01 (AB, J_{AB}=2, 4H-Ar); 7.09 (d, J=0.5, 2H₍₂₎Im); 7.20–7.32 (m, 6H_(m,p)-Ph); 7.86 (s, 2 OH); 9.43 (d, J=7.4, 2 NH). ¹³C NMR: 31.37, 32.10 (Me₃C); 32.23, 32.34 (Ar-CH₂-Ar); 32.74 (CH₂-Im); 34.27, 34.45 (Me₃C); 50.44 (CH₂-Ph); 52.50, 52.76 (C_αHis, OMe); 75.47 (OCH₂); 116.96, 125.37, 125.66, 125.92, 127.14, 127.30, 128.34, 129.14, 137.07 (3,5-Ar, 2,3,4,5,6-Ph, 2,4-Im); 126.77, 128.15, 132.69, 132.90, 137.13, 137.99, 142.62, 148.11, 150.34 (1,2,4,6-Ar, 5-Im, 1-Ph); 169.30 (CONH); 172.02 (COOMe). ES-MS (pos. mode): 1269.8 [12+Na]⁺; 646.4 [12+2 Na]²⁺/2; ES-MS (neg. mode): 1245.9 [12-H]⁻; 1268.0 [12+Na-2H]⁻. Anal. calcd for C₇₆H₉₀O₁₀N₆+H₂O (1264): C 72.12; H 7.32; N 6.64; O 13.91; found: C 71.98; H 7.37; N 6.68; O 13.93. Cobalt complexes: To a solution of ligand in CH₂Cl₂ was added solid CoCl₂ (3 equiv. for **7**, 2 equiv. for **12**). The solution turned blue immediately and was stirred under N₂ for 2 h. The excess of CoCl₂ which remained insoluble was filtered off and the complexes were directly precipitated by addition of hexane, giving quantitatively blue powders which were collected by filtration, then recrystallised from CH₂Cl₂/CH₃CN/EtOH mixture. Compound **13** (from 3 equiv. of CoCl₂ and 1 equiv. of **7**). IR (KBr): 1673.0 (CO(NH)), 1742.6 (CO(O)). UV (CH₂Cl₂): 274.5 (6200), 615 (320). ES-MS (pos. mode): 2127.9 [7+(CoCl₂)₂+Na⁺-H⁺]⁺, 2092.7 [7+(CoCl₂)₂+Na⁺-2H⁺-Cl⁻]⁺, 2069.7 [7+(CoCl₂)₂-Cl⁻]⁺, 2033.7 [7+(CoCl₂)₂-H⁺-2Cl⁻]⁺, 1939.8 [7+CoCl₂-Cl⁻]⁺, 1904.0 [7+CoCl₂-H⁺-2Cl⁻]⁺. Anal. calcd for C₁₀₈H₁₂₄O₁₆N₁₂ Co₂Cl₄·4H₂O (2177.94): C 59.51; H 6.06; N 7.71; Cl 6.51; Co 5.41; found: C 59.50; H 5.96; N 7.88; Cl 6.60; Co 5.04. Compound **14** (from 2 equiv. of CoCl₂ and 1 equiv. of **12**). IR (KBr): 1680.8 (CO(NH)), 1746.3 (CO(O)). UV (CH₂Cl₂): 281.5 (8000), 615 (185). ES-MS (pos. mode): 1340.4 [12+CoCl₂-Cl⁻]⁺, 1304.3 [12+CoCl₂-2Cl⁻-OH⁺]⁺, 1248.5 [12+H⁺]⁺. Anal. calcd for C₇₆H₉₀O₁₀N₆·CoCl₂·4 H₂O (1447.8): C 62.98; H 6.76; N 5.80; Cl 4.89; Co 4.06; found: C 63.19; H 6.49; N 5.96; Cl 5.04; Co 3.67.